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# CHROMOSOME NUMBERS IN LOTONONIS AND BUCHENROEDERA (FABACEAE—CROTALARIEAE)<sup>1</sup>

Ben-Erik Van Wyk and Anne Lise Schutte<sup>2</sup>

## ABSTRACT

Original chromosome counts for Buchenroedera (new generic report) and Lotononis (44 new specific reports) are presented. The most common somatic number in Lotononis, 18, was found in 29 species. Three species of Buchenroedera and nine species of Lotononis have 2n = 28. In Lotononis section Krebsia 2n = 28, 42, 56, and 84 were found in a closely related species group. This is the first report of a polyploid series in the Crotalarieae and includes the highest numbers recorded in the tribe. The chromosome numbers indicate anomalies in the existing sectional classification of Lotononis and may provide evidence for a more natural generic and infrageneric classification.

The genera Lotononis (DC.) Eckl. & Zeyh. and Buchenroedera Eckl. & Zeyh. are poorly known cytologically, with only six species of the former and none of the latter having been investigated previously. As part of an ongoing taxonomic study of these genera, chromosome counts were made for 47 species, representing almost the full range of variation in Lotononis (ca. 120 species centered in southern Africa, with a few extending into Asia) and Buchenroedera (ca. 16 species restricted to the eastern parts of southern Africa). The results are presented here, and their systematic significance in terms of an improved generic and infrageneric classification is discussed.

# MATERIALS AND METHODS

Mitotic counts were made from root tips of germinated seeds. Standard methods of pretreatment in hydroxyquinoline (0.02% mass/volume) and staining in lacto-propionic orcein were used. The duration of hydrolysis (1–8 minutes) and the concentration of HCl (0.2–0.5 N) proved to be important. The chromosomes are small (ca. 1–3  $\mu$ m long). Voucher specimens (listed in Table 1) are housed at the Rand Afrikaans University Herbarium (JRAU). A list of the species studied and voucher specimen details are given in the Appendix. Our

efforts to collect seeds have been rewarded by numerous rediscoveries of rare species and have provided a fairly representative sample of the two genera.

### RESULTS AND DISCUSSION

The results listed in Table 1 are arranged according to Duemmer's (1913) sectional classification. Where morphologically heterogenous sections of *Lotononis* have been subdivided into two or more groups, or where species have been moved to more appropriate positions, the reasons for doing so are given in the footnotes. The arrangement of species in Table 1 is aimed at facilitating the discussion that follows and is not intended as a formal infrageneric classification, but it nevertheless reflects major discontinuities and shows basic affinities

Several morphological characters provide links among the species of Lotononis with 2n = 28 and among those with 2n = 18. The latter are presently placed in various sections, indicating that Duemmer's infrageneric treatment is artificial; that the same chromosome number has evolved independently in several different groups seems unlikely. Section Krebsia, for example, presently comprises three distinct groups, two of which have obvious

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Table 1. Chromosome numbers in Lotononis and Buchenroedera. Species are arranged in sections following the treatment of Duemmer (1913), with some minor modifications that are explained in the footnotes. All known counts are included—those taken from the literature are preceded by an asterisk (\*).

Genera, Groups, and Species	Chromosome Number $(2n)$	Voucher or Reference
Buchenroedera Eckl. & Zeyh.		
B. lotononoides Scott Elliot	28	BVW 1966
B. meyeri Presl	28	BVW 1765
B. tenuifolia Eckl. & Zeyh. var. tenuifolia	28	BVW 1675
Lotononis (DC) Eckl. & Zeyh.		
Lotononis section Aulacinthus (E. Mey.) Benth.		
L. leucoclada (Schltr.) Duemmer	28	BVW 2430
L. gracilis (E. Mey.) Benth.	28	BVW 2250
otononis section Krebsia (Eckl. & Zeyh.) Benth.		
Part 1: Krebsia sensu stricto		
L. biflora (H. Bol.) Duemmer	± 84	BVW 1952
L. carnosa (Eckl. & Zeyh.) Benth.	84	BVW 1663
L. caerulescens (E. Mey.) B-E. van Wyk <sup>1</sup>	56	BVW 2483
L. cytisoides (E. Mey.) Benth.	28	BVW 1721
L. cytisoides (E. Mey.) Benth. aff.	56	BVW 1761
L. divaricata (Eckl. & Zeyh.) Benth.	56	BVW~2484
L. divaricata (Eckl. & Zeyh.) Benth. aff.	42	BVW 1666
L. trisegmentata Phill. var. robusta Phill. forma robusta	28	BVW 1917
L. trisegmentata Phill. var. robusta Phill. forma sericea Phill.	28	BVW 1956, 1958
Part 2: L. digitata group <sup>2</sup>	10	DIW 0041
L. digitata Harv.	18	BVW 2341
L. benthamiana Duemmer	18 18	BVW 2538
"L. magnifica" B-E. van Wyk ined.	10	BVW 2549
Part 3: L. transvaalensis group <sup>3</sup>	10	DI/W/ 10/0
L. transvaalensis Duemmer	18 18	BVW 1860 BVW 2504
L. procumbens H. Bol. 4	10	DVW 2304
otononis section Polylobium (Eckl. & Zeyh.) Benth.		
Part 1: Polylobium sensu stricto	90	DIZIV 9900
L. exstipulata L. Bol.	28 28	BVW 2280
*L. involucrata (Berg.) Benth.  *L. serpens (E. Mey.) Dahlgr. <sup>5</sup>	18	(Dahlgren, 1967) (Goldblatt, 1981b)
	10	(Goldblatt, 1901b)
Part 2: L. angolensis group <sup>6</sup> *L. angolensis Bak.	10	(Pth 1064)
*L. listii Polhill	18 18	(Byth, 1964) (Byth, 1964)
*L. bainesii Bak.	36	(Byth, 1964)
otononis section Telina (E. Mey.) Benth.	30	(Byth, 1904)
	90	DIVIV 9501
L. acuminata Eckl. & Zeyh.	28	BVW 2581
"L. repens" B-E. van Wyk ined.	28	BVW 2573
L. pungens Eckl. & Zeyh. <sup>7</sup> L. versicolor (E. Mey.) Benth. <sup>7</sup>	28 .28	BVW 1725 BVW 1386
	.20	DVW 1300
otononis section Oxydium Benth.*	10	DIZIV 0004
L. rostrata Benth.9	18	BVW 2324
L. rostrata aff.	18	BVW 2429
L. acutiflora Benth.	18	BVW 2544
L. oxyptera (E. Mey.) Benth.	18	BVW 2318
L. lenticula (E. Mey.) Benth.	18	BVW 2018
L. rabenaviana Dinter & Harms	18	BVW 2057

Table 1. Continued.

Genera, Groups, and Species	Chromosome Number $(2n)$	Voucher or Reference
Lotononis section Lipozygis (E. Mey.) Benth.		
Part 1: L. polycephala group <sup>10</sup>		
L. polycephala (E. Mey.) Benth.	18	BVW 2408
L. bolusii Duemmer	18	BVW 2443
"L. longicephala" B-E. van Wyk ined.	18	BVW 2241
Part 2: L. eriantha group <sup>11</sup>		
L. eriantha Benth.	18	ALS 383
L. foliosa H. Bol.	18	BVW 2607
L. lanceolata (E. Mey.) Benth.	18	BVW 1884
Lotononis section Leobordea (Del.) Benth.		
*L. platycarpa (Viv.) PicSerm.	18	(Goldblatt, 1981b)
Lotononis section Leptis (Eckl. & Zeyh.) Benth.		
Part 1: L. laxa group <sup>12</sup>		
L. laxa Eckl. & Żeyh.	18	BVW 2015
L. woodii H. Bol.	18	BVW 2608
L. macrosepala Conrath	18	BVW 2622
Part 2: L. brachyloba group <sup>13</sup>		
L. brachyloba (E. Mey.) Benth.	18	BVW 2244
"L. fruticoides" B-E. van Wyk ined.	18	BVW 2020
L. leptoloba H. Bol.	18	ALS 276
L. maximilianii Schltr. (cleistogamous)	18	ALS 271
L. maximilianii (chasmogamous)	18	ALS 282
Part 3: L. calycina group <sup>14</sup>		
L. calycina (E. Mey.) Benth.	18	BVW 2621
L. sericoflora Duemmer	18	BVW 1899
L. humifusa Benth.	18	BVW 1700
L. mucronata Conrath aff.	18	BVW 2619
"L. curvicarpa" B-E. van Wyk ined.	18	BVW 2725

- <sup>1</sup> Better known as Lebeckia microphylla E. Mey.
- <sup>2</sup> Species added to section Krebsia by Harvey (1862) and Duemmer (1913).
- <sup>3</sup> Species added to Krebsia by Duemmer (1913).
- <sup>4</sup> Position in section *Polylobium* was based on a superficial characterization.
- <sup>5</sup> An anomalous species. Dahlgren (1964) suggested similarities with L. involucrata.
- <sup>6</sup> Species added to section *Polylobium* by Baker (1871) and related species.
- <sup>7</sup> Superficially similar to L. laxa and previously associated with the section Leptis.
- <sup>8</sup> This section was referred to the genus Crotalaria by Duemmer (1913).
- <sup>9</sup> Better known as L. micrantha (E. Mey.) Benth.
- <sup>10</sup> A distinct group of *Lipozygis* with indehiscent, wind-dispersed fruit.
- 11 A distinct group of pyrophytes from grassland areas of the eastern parts of southern Africa.
- <sup>12</sup> Perennial herbs with acute keel petals as in section Oxydium.
- <sup>13</sup> Annuals with acute keel petals as in section Oxydium.
- <sup>14</sup> Annuals and perennials with obtuse keel petals as in the L. eriantha group of section Lipozygis.

affinities elsewhere in the genus. The woody habit of *L. digitata* and *L. transvaalensis* was used to place them in *Krebsia*, but both are morphologically very similar to various species of section *Leptis*. Another example is section *Polylobium*; *Lotononis umbellata* and its allies are closely related to section *Aulacinthus* and perhaps not distinct

from it at the sectional level. The *L. angolensis* group is quite different from other species of section *Polylobium* and its position in this section is unsatisfactorily artificial.

Two separate phylogenetic lines with base numbers of x = 9 and x = 7 are suggested, and further research will show if other evidence supports such

a dichotomy in the genus. Not a single count of 2n = 16 or 32 has been made, so that a base number of 8, which is common in some of the other genera, so far appears to be totally absent in *Lotonomis*.

At the generic level, the data also give some indications of affinity. Buchenroedera is so closely related to Lotononis (especially to section Krebsia) that its generic status has been questioned (Polhill, 1976, 1981). The shared chromosome number of 2n = 28 (and presumably a base number of 7) agrees with chemical evidence (Van Wyk & Verdoorn, 1988) that Buchenroedera is perhaps best considered a section of Lotononis.

The remarkable similarities between species of Crotalaria and Lotononis have caused confusion in past taxonomic treatments. For example, most species of Lotononis section Oxydium were transferred to Crotalaria by Duemmer (1913). The presence of macrocyclic pyrrolizidine alkaloids in both genera (Van Wyk & Verdoorn, in prep.) indeed indicates that Lotononis is more closely related to Crotalaria than to other genera of the tribe, all of which seem to contain only quinolizedine alkaloids. Crotalaria, however, have 2n = 16, 32, or rarely 14 (Goldblatt, 1981a), while those species of Lotononis that closely resemble Crotalaria (section Oxydium and some groups of Leptis) all have 2n = 18. The morphological distinction between Lotononis and Crotalaria (Polhill, 1968) is therefore strongly supported by the data at hand.

Some of the woody species of Lotononis (sections Aulacinthus and Krebsia) are very similar to species of Lebeckia. Lotononis caerulescens (E. Mey.) B-E. van Wyk, for example, has until recently been known as Lebeckia microphylla E. Mey., but morphological and chemical evidence (Van Wyk, 1988; Van Wyk & Verdoorn, 1988) clearly showed it to be misplaced in Lebeckia. The sections Aulacinthus and Krebsia sensu stricto have 2n = 28, 42, 56, and 84, while four counts of 2n = 18 are known for Lebeckia (Dahlgren, 1967). Here again, the cytological data agree with the morphological distinction between Lotononis and Lebeckia. Lotononis angolensis and related species (section *Polylobium*) are chemically similar to Lebeckia and also have the same chromosome number. Morphological characters such as the zygomorphic calyx and dimorphic stipules, however, are typical of Lotononis.

Not a single count of 2n = 14 is known for *Lotononis*; so it seems to be cytologically different from the genus *Pearsonia*. The only available count for the latter genus was by Frahm-Leliveld (1969),

who reported 2n = 14 for P. flava (Bak. f.) Polhill. The species of Pearsonia are similar to Lotononis except for their highly modified flowers (Polhill, 1973), and the shared chromosome base number of x = 7 may indeed indicate a common ancestry.

From a phylogenetic point of view, the different base numbers in Lotononis suggest interesting questions about generic relationships in the Crotalarieae. The base number of the tribe is almost certainly x = 9 (Goldblatt, 1981a), and 2n = 18in some species of Lotononis is presumably the ancestral condition. The only way to achieve 2n= 28 (if *Lotononis* is monophyletic) is to postulate descending an euploidy from n = 9 to 8 and 7 and subsequent polyploidy. Since 2n = 16 and 14appear to be totally absent in Lotononis, it may be argued that Crotalaria and Pearsonia form part of the lineage that gave rise to the group of species with 2n = 28, 42, 56, and 84. If Lotononis proves to be polyphyletic, this possibility can be seriously considered, but the generic characters of the current concept of Lotononis are present in at least some species of each major group. Although there are marked phenetic similarities linking all the major groups, Lotononis as a whole is not monothetic. It is defined by combinations of apomorphic tendencies, such as single stipules, suffrutescent or herbaceous habit, absence of bracteoles, fusion of the lateral calyx lobes, verrucose upper suture of the fruit, tuberculate testa, elongated funicles, flower dimorphism associated with cleistogamy, ability to produce HCN, and presence of macrocyclic pyrrolizidine alkaloids. There is not a single apomorphy known to us that would unambiguously support monophyly. A possible solution would be to separate the lineage with 2n = 28from the one with 2n = 18 and to split the latter into several smaller groups. Despite conflicting character information, there are some indications from the morphology that the geographically widespread and generally herbaceous 2n = 18 lineage is more primitive than the predominantly woody and essentially southern African 2n = 28 lineage.

In a tribal context, the occurrence of polyploidy in Lotononis (section Krebsia) is of some interest. Polyploidy and high chromosome numbers are typical of the Genisteae but have never been reported from any genus of the Crotalarieae (Goldblatt, 1981a). It is also noteworthy that polyploidy should occur in an essentially woody group (previously considered to be one of the basal groups of Lotononis) and not in the supposedly more derived herbaceous groups. Unlike the situation in the other large genera of the Crotalarieae (Áspalathus and

to some extent *Crotalaria*), there is no direct evidence of aneuploidy, although it must have played a significant role in the phylogeny of *Lotononis*.

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# APPENDIX

List of species, collection data, and voucher specimen details of the material used for chromosome counts. Voucher specimen numbers refer to our own collections (abbreviated as *BVW* and *ALS*) and are all housed in the Rand Afrikaans University Herbarium (JRAU). Authorities for names are given in Table 1.

Buchenroedera lotononoides: Loteni, Natal, BVW 1966. B. meyeri: Mhlahlane, Transkei, BVW 1765. B. tenuifolia var. tenuifolia: Queenstown, E Cape, BVW 1675.

Lotononis acuminata: Humansdorp district, S Cape, BVW 2581. L. acutiflora: Khamiesberg, Cape, BVW 2544. L. benthamiana: Springbok district, Cape, BVW 2538. L. biflora: Loteni, Natal, BVW 1952. L. bolusii: Piquetberg, Cape, BVW 2443. L. brachyloba: Ceres, Cape, BVW 2244. L. caerulescens: Cradock, E Cape, BVW 2483. L. calycina: Bethal, Transvaal, BVW 2621. L. carnosa: Queenstown, E Cape, BVW 1663. "L. curvicarpa" (ined.): Devon, Transvaal, BVW 2725. L. cytisoides: Winterberg, E Cape, BVW 1721. L. cytisoides aff.: Mhlahlane, Transkei, BVW 1761. L. digitata: Garies, Cape, BVW 2341. L. divaricata: Swagershoek Pass, E Cape, BVW 2484. L. divaricata aff.: Queenstown, E Cape, BVW 1666. L. eriantha: Roodepoort, Transvaal, ALS 383. L. exstipulata: Ceres district, Cape, BVW 2280. L. foliosa: Johannesburg. Transvaal, BVW 2607. "L. fruticoides" (ined.): Graaff Reinet district, Cape, BVW 2020. L. gracilis: Ceres, Cape, BVW 2250. L. humifusa: Grahamstown district, E Cape, BVW 1700. L. lanceolata: Dullstroom, Transvaal, BVW 1884. L. laxa: Colesberg, Cape, BVW 2015. L. lenticula: Colesberg, Cape, BVW 2018. L. leptoloba: Nieuwoudtville, Cape, ALS 276. L. leucoclada: Clanwilliam, Cape, BVW 2430. "L. longicephala" (ined.): Touw's River, Cape, BVW 2241. L. macrosepala: Bethal district, Transvaal, BVW 2622. "L. magnifica" (ined.): Khamiesberg, Cape, BVW 2549. L. maximiliani: Nieuwoudtville, Cape, ALS 271 (cleistogamous form), ALS 282 (chasmogamous form). L. mucronata aff.: Ermelo district, Transvaal, BVW 2619. L. oxyptera: Citrusdal, Cape, BVW 2318. L. polycephala: Khamiesberg, Cape, BVW 2408. L. procumbens: Volksrust district, Natal, BVW 2504. L. pungens: Tarkastad district, E Cape, BVW 1725. L. rabenaviana: Beaufort West district, Cape, BVW 2057. "L. repens" (ined.): Outeniqua Pass, S Cape, BVW 2573. L. rostrata: Citrusdal, Cape, BVW 2324. L. rostrata aff.: Klawer, Cape, BVW 2429. L. sericoflora: Harrismith, Orange Free State, BVW 1899. L. transvaalensis: Nelspruit, E Transvaal, BVW 1860. L. trisegmentata var. robusta forma robusta: Clarens, Orange Free State, BVW 1917. L. trisegmentata var. robusta forma sericea: Loteni, Natal, BVW 1956; Sani Pass, Natal, BVW 1958. L. versicolor: Beaufort West district, Cape, BVW 1386. L. woodii: Wakkerstroom district, Natal, BVW 2608.